## Prebiotic Activity of Ambon Banana (*Musa acuminata* (AAA Group) 'Ambon') Peel Starch Against *Lactobacillus. acidophilus* and *Escherichia coli* In Vitro

### Bertha Rusdi<sup>\*</sup>, Umi Yuniarni

Apothecary Program, Faculty of Mathematics and Sciences, Universitas Islam Bandung, Bandung, Jawa Barat, Indonesia

\*Corresponding Author Email: bertha.rusdi@unisba.ac.id

Submit: November 16<sup>th</sup>, 2023

Revised: December 2<sup>nd</sup>, 2023

Accept: December 16th, 2023

Copyright © 2023 Universitas Pakuan

FITOFARMAKA: Jurnal Ilmiah Farmasi is licensed under a Creative Commons Attribution-ShareAlike 4.0 International License

## ABSTRACT

Prebiotics are compounds with the ability to specifically enhance the population of advantageous bacteria in the gastrointestinal tract. Starch is a polysaccharide which has prebiotic activity. Agricultural waste like banana peel contains prebiotic polysaccharides including starch. The prebiotic effect of starch from many varieties of banana peel has been explored by researchers. Although Ambon banana is a variety that is commonly consumed in Indonesia, the prebiotic activity of its peel, particularly the peel starch, has not been studied yet. Thus, this study aims to research the prebiotic activity of Ambon banana peel starch against probiotic bacteria of the Lactobacillus acidophilus and the opportunistic pathogen bacteria of Eschericia coli. In this research, starch was extracted from banana peel (var. pisang Ambon) and prebiotic activity of the starch was tested on L. acidophilus and E. coli. The number of bacteria was calculated at 0 and 24 hours of incubation using plate count methods. The result showed that at the concentration of 1% w/v, Ambon banana peel starch increases the number of L. acidophilus while inhibiting the growth of E. coli. The L. acidophilus culture in the starch-containing media had SCFAs (acetic, butyric and propionic acid) that were known to have good impact to human health.

Keywords: Prebiotic; ambon banana peel; starch; L. acidophilus.

## INTRODUCTION

Prebiotics are compounds that cannot be digested but can be used by beneficial bacteria in the digestive tract so that the growth of these bacteria increases (Davani-Davari et al., 2019). The increase of beneficial bacteria in the digestive tract results in good benefits for the host's health (humans), including preventing the emergence of several diseases such as diabetes, cancer, and cardiovascular diseases (Oniszczuk et al., 2021). The magnitude of the health benefits due to the consumption of prebiotics has increased the demand for products containing prebiotics (Farias et al., 2019). Numerous investigations have been undertaken to explore cost-effective origins of prebiotics. Agricultural wastes such as rice straw, fruit peels, and corncobs are known to be the potential source of cheap prebiotics (Le & Yang, 2019; Samanta et al., 2015). Polysaccharides or oligosaccharides contained in agricultural waste such as pectin, cellulose, starch, inulin, fructooligosaccharides (FOS), pectic-oligosaccharides, xylo-oligosaccharides have prebiotic effects (Bamigbade et al., 2022). Banana peel is one of the agricultural wastes which is rich in carbohydrates including starch (Aquino et al., 2016; Khamsucharit et al., 2018; Romelle et al., 2016). Bananas are one of the fruits with high production and consumption in Indonesia, thus banana peels are available in large amount (Wisnubroto, 2021).

The prebiotic effect of banana peels from several varieties have been studied. Raw "saba/kepok" banana peel powder can increase the number of Lactobacillus bacteria in vitro. A similar effect was also found in "silver bluggoe" banana peel powder (white costa banana) (Powthong et al., 2020). Banana peel of Musa sapientum ("Kluai Namwa") variety contains resistant starch which shows a prebiotic effect against *Bacteroides vulgatus* ATCC 8482, *Bifidobacterium longum* ATCC BAA-999, *Lactobacilus casei* subsp. rhamnosus TISTR 047, and *Lactobacilus acidophilus* TISTR 450 (Jaiturong et al., 2020a).

Ambon banana (Musa acuminata Triploid AAA) is a banana variety that is widely consumed in Indonesia, the fruit flesh is consumed directly or processed into other foods such as chips and cakes, but its peel is rarely used. Ambon banana contains starch, a prebiotic polysaccharide (Musita, 2012). Ambon bananas with peels increased the growth of probiotic lactic acid bacteria at concentrations of 7.5 and 10% (Handayani, 2021). However, the prebiotic effect of starch from Ambon banana peels has not been specifically studied, thus this study aims to research the prebiotic activity of Ambon banana peel starch against probiotic bacteria of the L. acidophilus and opportunistic pathogen bacteria of the E. coli. The prebiotic effect was indicated by increasing the number of L. acidophilus while decreasing the number of E. coli. The results of this study are expected to be used as a basis for further research for the development of synbiotic products derived from Ambon banana peel waste as a supplement to treat diseases related to the immune system or metabolic disorders.

# METHODS

## Equipments

High performance liquid chromatography (dionex ultimate 3000 Thermo scientific), pHmeter (SevenCompactTM pH/Ion S 220, Metler-Toledo), Spectrophotometer (UV-1800, Shimadzu), oven (UNB 200, Memmert), blender (HR 2116, Phillips).

# Materials

Ambon banana peels were collected from one local cake shop in Bandung, West Java. The variety of banana was determined at Sekolah Ilmu dan Teknologi Hayati (SITH), Institut Teknologi Bandung (ITB), NaOH (technical grade, Bratchem), Lactobacilus acidophilus (ATCC 4346), Escherichia coli (ATCC 25922), nutrient broth (NB, Merck), De Man, Rogosa and Sharpe (MRSA, Oxoid), nutrient agar (NA, Merck).

# Methods

## Starch Extraction

Starch was extracted from banana peels using the procedure described in the study conducted by Uraipan et al. (2014). Banana peels were cut into thin slices and then dried at 55 °C for 7 hours. After drying, starch extraction was carried out using NaOH (technical grade, Bratchem) and water in a ratio of 1:1. Then the slurry was filtered. The starch precipitate was rinsed several times with distilled water. Thereafter the starch was dried at 65 °C for 15 hours in an oven.

# **Total Carbohydrate Determination**

Phenol-sulfuric The method was employed to determine the total carbohydrate concentration in the starch (Nielsen, 2017). Briefly, A100 mg/L glucose stock solution was prepared and serial dilutions were then performed by adding the glucose stock solution to distilled water. Ambon banana peel starch (40 mg) was dissolved in 10 mL of distilled water, and 1 mL of the resulting solution was mixed with 1 mL of water to create duplicates. To these solutions,  $500 \,\mu L \,(0.05 \,m L)$  of  $80 \,\%$  phenol was added and vortexed. Subsequently, 5 mL of sulfuric acid was added, and the tubes were vortexed and left in a water bath at 25 °C for 10 minutes. The absorbance of each solution was measured at a wavelength of 490 nm. The absorbance of Ambon banana peel starch was

substituted into the regression equation of the standard curve to quantify the total carbohydrate content.

#### **Bacterial Rejuvenation**

The prebiotic activity of starch was tested against Lactobacilus acidophilus (ATCC 4346), a representative of probiotic bacteria and Escherichia coli (ATCC 25922), as a representative of the unfavorable microflora which were obtained from Microbiology Laboratory of Pharmacy Department, Universitas Islam Bandung. De Man, Rogosa and Sharpe Broth (MRSB, Oxoid) without glucose was used as a medium for the growth of L. acidophilus and nutrient broth (NB, Merck) for the growth of E. coli. All bacteria were maintained at 37 °C and re-cultured in fresh media every 16-24 hours.

### **Prebiotic Activity Test**

The media containing 1 % (w/v) of Ambon banana peel starch was prepared. The media without glucose, media with 1 % (w/v) of inulin and media with 1 % (w/v) glucose were used as control. About 10 mL of each media was added with *L. acidophilus* and *E. coli* culture suspension to the final bacteria number of 1 x  $10^8$  CFU/mL. The cultures were incubated at  $37^{\circ}$ C for 24 hours. The number of bacteria in the samples at 0 and 24 hours of incubation was determined using the plate count method. The media De Man, Rogosa and Sharpe agar (MRSA, Oxoid) was used for counting *L. acidophilus* and nutrient agar (NA, Merck) for counting *E. coli*. The number of bacterial colonies was calculated as the CFU/mL value. The equation 1 was used in calculating the prebiotic index (PI) of the starch (Phirom-on & Apiraksakorn, 2021).

#### SCFAs Analysis and pH Measurements

The SCFAs analysis was conducted at the Laboratory of Pusat Studi Pengembangan Sediaan Farmasi Fakultas Farmasi Universitas Padjadjaran. The concentration of short fatty acid (SCFA) in the form of butyrate, propionate and acetate were measured using high performance chromatography liquid under isocratic conditions using C<sub>18</sub> column (Universal Fortis) and measured at a Column  $C_{18}$  15 cm, wave length 210 nm, mobile phase 5 % acetonitrile in 0.012 N H<sub>2</sub>SO<sub>4</sub>, flow rate 0.5 mL/min., and injection volume 20 µL. The pH value of bacterial culture solution containing banana peel starch was measured using a pH meter at 0, 24, 48 and 72 hours of incubation.

#### **Statistical Analysis**

The difference of bacteria number between groups were analyzed statistically using ANOVA followed with Tukey Test. The difference of *L. acidophilus* and *E. coli* number in media with Ambon banana peel starch was analyzed using T-Test.

$$\mathbf{PI} = \left[ \frac{(P_p^{24} - P_p^0) - (P_{GF}^{24} - P_{GF}^0)}{(P_G^{24} - P_G^0) - (P_{GF}^{24} - P_{GF}^0)} \right] - \left[ \frac{(E_p^{24} - E_p^0) - (E_{GF}^{24} - E_{GF}^0)}{(E_G^{24} - E_G^0) - (E_{GF}^{24} - E_{GF}^0)} \right]$$
(1)

Where:

 $\begin{array}{ll} P_p^0, P_p^{24} &= \text{CFU for probiotic } (L. acidophilus) \text{ at } 0 \text{ and } 24 \text{ h in media enriched with starch} \\ P_G^0, P_G^{24} &= \text{CFU for probiotic } (L. acidophilus) \text{ at } 0 \text{ and } 24 \text{ h in glucose-free media} \\ P_G^0, P_G^{24} &= \text{CFU for probiotic } (L. acidophilus) \text{ at } 0 \text{ and } 24 \text{ h in media enriched with} \quad \text{glucose} \\ E_p^0, E_p^{24} &= \text{CFU for pathogen } (E. coli) \text{ at } 0 \text{ and } 24 \text{ h in media enriched with starch} \\ E_{GF}^0, E_{GF}^{24} &= \text{CFU for pathogen } (E. coli) \text{ at } 0 \text{ and } 24 \text{ h in media enriched with starch} \\ E_{GF}^0, E_{GF}^{24} &= \text{CFU for pathogen } (E. coli) \text{ at } 0 \text{ and } 24 \text{ h in glucose-free media} \\ E_G^0, E_G^{24} &= \text{CFU for pathogen } (E. coli) \text{ at } 0 \text{ and } 24 \text{ h in glucose-free media} \\ E_G^0, E_G^{24} &= \text{CFU for pathogen } (E. coli) \text{ at } 0 \text{ and } 24 \text{ h in media enriched with glucose} \end{array}$ 

### **RESULTS AND DISCUSSION**

The determination of the banana variety being used in this research by Sekolah Ilmu dan Teknologi Hayati (SITH), Institut Teknologi Bandung (ITB) showed that it was Musa acuminata (AAA Group) 'Ambon' (number of certificate 5214/IT1.C11.2/TA.00/2022). The yield of starch from fresh Ambon banana peel was 0.67 %. While the total carbohydrate content in the starch was 44.26 % w/w. The effect of starch Ambon banana peel as a carbon source was tested on the growth of the probiotic bacteria, L. acidhopilus and the opportunist microbiota E. coli. In this test, glucose is employed as the positive control, as it is known to be a carbon source that can be utilized by various types of bacteria (Sánchez-Clemente et al., 2020). Inulin is used as a reference because it is a known prebiotic compound that can enhance the growth of beneficial bacteria (Pandey et al., 2015). Bacteria grown on media without glucose are used as the negative control. It is hypothesized that bacteria cultivated in this medium will experience slower growth compared to bacteria grown in media containing a carbon source. The results are presented in Table 1.

ANOVA analysis results showed that the number of *L. Acidophilus* that grew in media containing Ambon banana peel starch were significantly higher than the bacteria in media with inulin or without carbon source. The number of L. Acidophilus in media with Ambon banana peel starch was not significantly different to glucose. Meanwhile the E. coli number in media with Ambon banana peel starch was significantly lower than in media media without carbon source and/or inulin. Furthermore, the statistical analysis (T-test) of bacterial count in cultures of L. acidophilus and E. coli treated with Ambon banana peel starch, indicates that the quantity of L. acidophilus bacteria was significantly higher than that of E. coli. Thus, it can be concluded that Ambon banana peel starch has prebiotic activity because it can increase the growth of beneficial bacteria (L. acidophilus) and suppress the growth of opportunistic bacteria (E. coli). While, the PI of Ambon banana peel starch was 0.21. This result supports findings in the previous study which reported that banana peels had a prebiotic effect on lactic acid bacteria (LAB (Zahid et al., 2021). However, the PI of Ambon banana peel starch was lower than reported by previous research which used unripe banana of Kluai Namwa Luang (pisang awak) variety (Jaiturong et al., 2020b). This might be due to the starch used in this study was from ripe bananas while the ripe banana was known to have lower content of prebiotic in the form of resistant starch than unripe banana (Li et al., 2020; Phillips et al., 2021).

Table 1.	Effect of Ambon Banana Peel Starch on The Growth of L. acidophilus and E. coli After 24
	Hours of Incubation

	Number of Bacteria (Log CFU/mL)					
Bootorio	0.b	24 h				
Dacteria	(for all groups)	Without carbon source	Glucose	Inulin	Ambon banana peel starch	
L. acidophilus	$8\pm0$	$6.62 \pm 0.24$	$8.13 \pm 0.26^*$	$7 \pm 0.52$	$9.11 \pm 0.71*$	
E. coli	$8\pm0$	$11.86 \pm 0.78$	$8.34 \pm 0.48*$	$10.57\pm0.10$	$6.81 \pm 0.22*$	

Data are in mean  $\pm$  SD (n=3). \*) is significantly different compared to bacteria in media without carbon source/inulin (p<0.5)



Figure 1. The pH of L. acidophilus culture that enriched with Ambon banana peel starch

Inulin was reported as a potential prebiotic and could selectively increase *L*. *acidophilus* (Carlson et al., 2018; Setiarto et al., 2017; Zarinah et al., 2018). However, in this study, inulin showed no prebiotic effect, and its PI was - 0.11. The negative result of inulin prebiotic activity was also reported in previous studies (Adebola et al., 2014; Bedani et al., 2013).

*Lactobacilli* converts polysaccharides into SCFA (Zhao et al., 2021). Increased levels of SCFA might be indicated by a decrease in the pH of the bacterial culture solution. In 72 hours, pH of *L. acidophilus* culture that contains Ambon banana peel starch was declining (Figure 1.). Furthermore, the concentration of SCFA in the culture after 24 hours incubation was determined (Table 2).

Table 2.	Concentratio	n of	SCFA	in	L.
	acidophilus	culture	that	enric	hed
	with Ambon	banana	peel sta	arch	

SCFA	Conc	Concentration (µg/mL)			
	1	2	mean		
Acetic acid	8.03	10.63	9.33		
Butyric acid	5.69	6.06	5.88		
Propionic acid	0.81	0.77	0.79		

Acetic acid was the highest SCFA contained in the *L. acidophilus* culture media enriched with 1 % b/v Ambon banana peel

starch followed by butyric acid and lastly propionic acid. This trend was similar to SCFAs content of human faecal slurry incubated with banana flesh powder (Musa acuminata Colla) 0.2 % b/v (Tian et al., 2020). Acetic acid is a SCFA known to play a role in controlling body weight and insulin sensitivity (Hernández et al., 2019). Butyric acid, which is a source of energy for intestinal cells, can improve intestinal peristalsis and has anti-inflammatory activity (Pituch et al., 2013; Xu et al., 2021; Załęski et al., 2013). While propionic acid is associated with lowering blood lipid levels and preventing cancer (Hosseini et al., 2011). This finding suggests that Ambon banana peel starch might exert a beneficial effect on health.

#### CONCLUSIONS

Ambon banana peel starch had a prebiotic effect in vitro which can be seen from their ability to increase the number of *L. acidophilus* and suppress the growth of *E. coli*. Acetic acid, butyric acid and propionic were detected in the *L. acidophilus* culture which grew in Ambon banana peel starch containing media. The presence of those SCFAs were also indicated by pH reduction over 72 hours period of incubation. Further study in the animal model is needed to confirm the prebiotic effect of Ambon banana peel starch.

## ACKNOWLEDGE

This research was funded by Lembaga Penelitian dan Pengabdian Kepada Masyarakat (LPPM), Universitas Islam Bandung (Contract number: 068/B.04/LPPM/XII/2021). The authors would like to thank Dieni Mardliani, Faqih Radina, Ermawati and Winda Amelya who have aided in conducting experiments in the laboratory and Hendra Gunawan for his assistance in conducting statistical analysis. We also thank to Laboratory of Pusat Studi Pengembangan Sediaan Farmasi Fakultas Farmasi Universitas Padjadjaran for their assistance in analysing the SCFAs content in the bacteria cultures.

## BIBLIOGRAPHY

- Adebola, O. O., Corcoran, O., & Morgan, W. A. (2014). Synbiotics: the impact of potential prebiotics inulin, lactulose and lactobionic acid on the survival and growth of lactobacilli probiotics. *Journal of Functional Foods*, 10, 75–84. https://doi.org/https://doi.org/10.1016/j.jff. 2014.05.010
- Aquino, C. F., Salomão, L. C. C., Ribeiro, S. M.
  R., Siqueria, D. L. D. E., & Cecon, P. R.
  (2016). Carbohydrates, phenolic compounds and antioxidant activity in pulp and peel of 15 banana cultivars. *Revista Brasileira de Fruticultura*, 38.
- Bamigbade, G. B., Subhash, A. J., Kamal-Eldin,
  A., Nyström, L., & Ayyash, M. (2022).
  An Updated Review on Prebiotics: Insights on Potentials of Food Seeds
  Waste as Source of Potential Prebiotics. In *Molecules* (Vol. 27, Issue 18). https://doi.org/10.3390/molecules271859 47
- Bedani, R., Rossi, E. A., & Saad, S. M. I. (2013).
  Impact of inulin and okara on Lactobacillus acidophilus La-5 and Bifidobacterium animalis Bb-12 viability in a fermented soy product and probiotic survival under in vitro simulated gastrointestinal conditions. Food

*Microbiology*, *34*(2), 382–389. https://doi.org/10.1016/j.fm.2013.01.012

- Carlson, J. L., Erickson, J. M., Lloyd, B. B., & Slavin, J. L. (2018). Health Effects and Sources of Prebiotic Dietary Fiber. *Current Developments in Nutrition*, 2(3), nzy005. https://doi.org/10.1093/cdn/nzy005
- Davani-Davari, D., Negahdaripour, M., Karimzadeh, I., Seifan, M., Mohkam, M., Masoumi, S. J., Berenjian, A., & Ghasemi, Y. (2019). Prebiotics: Definition, types, sources, mechanisms, and clinical applications. *Foods*, 8(3), 1– 27. https://doi.org/10.3390/foods8030092
- Farias, D. de P., de Araújo, F. F., Neri-Numa, I. A., & Pastore, G. M. (2019). Prebiotics: Trends in food, health and technological applications. *Trends in Food Science & Technology*, 93, 23–35. https://doi.org/https://doi.org/10.1016/j.tifs .2019.09.004
- Handayani, I. N. A. (2021). Potensi Yogurt Dengan Penambahan Lactobacillus casei dan Tepung Pisang Ambon Dalam Menghambat Patogen. *JITIPARI*, 6(2), 1– 13.

https://ejurnal.unisri.ac.id/index.php/jtpr/a rticle/download/5008/pdf

- Hernández, M. A. G., Canfora, E. E., Jocken, J. W. E., & Blaak, E. E. (2019). The Short-Chain Fatty Acid Acetate in Body Weight Control and Insulin Sensitivity. *Nutrients*, *11*(8). https://doi.org/10.3390/nu11081943
- Hosseini, E., Grootaert, C., Verstraete, W., & Van de Wiele, T. (2011). Propionate as a health-promoting microbial metabolite in the human gut. *Nutrition Reviews*, 69(5), 245–258. https://doi.org/10.1111/j.1753-4887.2011.00388.x
- Jaiturong, P., Laosirisathian, N., Sirithunyalug, B., Eitssayeam, S., Sirilun, S., Chaiyana, W., & Sirithunyalug, J. (2020a).
  Physicochemical and prebiotic properties of resistant starch from Musa sapientum Linn., ABB group, cv. Kluai Namwa

Luang. *Heliyon*, 6(12), e05789. https://doi.org/10.1016/j.heliyon.2020.e05 789

- Jaiturong, P., Laosirisathian, N., Sirithunyalug, B., Eitssayeam, S., Sirilun, S., Chaiyana, W., & Sirithunyalug, J. (2020b). Potential of Musa sapientum Linn. for digestive function promotion by supporting Lactobacillus sp. *Heliyon*, *6*(10), e05247. https://doi.org/https://doi.org/10.1016/j.hel iyon.2020.e05247
- Khamsucharit, P., Laohaphatanalert, K., Gavinlertvatana, P., Sriroth, K., & Sangseethong, K. (2018). Characterization of pectin extracted from banana peels of different varieties. *Food Science and Biotechnology*, 27(3), 623– 629.
- Le, B., & Yang, S. H. (2019). Production of prebiotic xylooligosaccharide from aqueous ammonia-pretreated rice straw by β-xylosidase of Weissella cibaria. *Journal of Applied Microbiology*, 126(6), 1861–1868.
- Li, M.-C., Chou, C.-F., Hsu, S.-C., & Lin, J.-S. (2020). Physicochemical characteristics and resistant starch of different varieties of banana from Taiwan. *International Journal of Food Properties*, 23(1), 1168– 1175.

https://doi.org/10.1080/10942912.2020.1 788077

- Musita, N. (2012). Kajian Kandungan dan Karakterisik Pati Resisten dari Berbagai Varietas Pisang. *Jurnal Teknologi & Industri Hasil Pertanian*, 14(1), 68–79. http://dx.doi.org/10.28959/jdpi.v23i1.557
- Nielsen, S. S. (2017). Total Carbohydrate by Phenol-Sulfuric Acid Method BT - Food Analysis Laboratory Manual (S. S. Nielsen (ed.); pp. 137–141). Springer International Publishing. https://doi.org/10.1007/978-3-319-44127-6\_14
- Oniszczuk, A., Oniszczuk, T., Gancarz, M., & Szymańska, J. (2021). Role of Gut Microbiota, Probiotics and Prebiotics in the Cardiovascular Diseases. In

*Molecules* (Vol. 26, Issue 4). https://doi.org/10.3390/molecules260411 72

- Pandey, K. R., Naik, S. R., & Vakil, B. V. (2015). Probiotics, prebiotics and synbiotics- a review. *Journal of Food Science and Technology*, 52(12), 7577– 7587. https://doi.org/10.1007/s13197-015-1921-1
- Phillips, K. M., McGinty, R. C., Couture, G., Pehrsson, P. R., McKillop, K., & Fukagawa, N. K. (2021). Dietary fiber, starch, and sugars in bananas at different stages of ripeness in the retail market. *PloS One*, 16(7), e0253366. https://doi.org/10.1371/journal.pone.0253 366
- Phirom-on, K., & Apiraksakorn, J. (2021). Development of cellulose-based prebiotic fiber from banana peel by enzymatic hydrolysis. *Food Bioscience*, *41*, 101083. https://doi.org/10.1016/j.fbio.2021.10108 3
- Pituch, A., Walkowiak, J., & Banaszkiewicz, A. (2013). Butyric acid in functional constipation. *Przeglad Gastroenterologiczny*, 8(5), 295–298. https://doi.org/10.5114/pg.2013.38731
- Powthong, P., Jantrapanukorn, B., Suntornthiticharoen, P., & Laohaphatanalert, K. (2020). Study of prebiotic properties of selected banana species in Thailand. *Journal of Food Science and Technology*, *57*(7), 2490– 2500. https://doi.org/10.1007/s13197-020-04284-x
- Romelle, F. D., Rani, A., & Manohar, R. S. (2016). Chemical composition of some selected fruit peels. *European Journal of Food Science and Technology*, 4(4), 12– 21.
- Samanta, A. K., Jayapal, N., Jayaram, C., Roy, S., Kolte, A. P., Senani, S., & Sridhar, M. (2015). Xylooligosaccharides as prebiotics from agricultural by-products: production and applications. *Bioactive Carbohydrates and Dietary Fibre*, 5(1), 62–71.

Sánchez-Clemente, R., Guijo, M. I., Nogales, J., & Blasco, R. (2020). Carbon Source Influence on Extracellular pH Changes along Bacterial Cell-Growth. *Genes*, *11*(11).

https://doi.org/10.3390/genes11111292

- Setiarto, R. H. B., Widhyastuti, N., Saskiawan, I., & Safitri, R. M. (2017). Pengaruh Variasi Konsentrasi Inulin Pada Proses Fermentasi Oleh L. Acidophilus, L. Bulgaricus Dan S. Thermophillus - (the Inulin Variation Concentration Effect in Fermentation Using L. Acidophilus, L. **Bulgaricus** and S. Thermophilus). Biopropal Industri, 8(1), 1–17. https://doi.org/10.36974/jbi.v8i1.1669
- Tian, D. D., Xu, X. Q., Peng, Q., Zhang, Y. W., Zhang, P. B., Qiao, Y., & Shi, B. (2020). Effects of banana powder (Musa acuminata Colla) on the composition of human fecal microbiota and metabolic output using in vitro fermentation. *Journal of Food Science*, 85(8), 2554– 2564. https://doi.org/10.1111/1750-3841.15324
- Uraipan, S., Brigidi, P., & Hongpattarakere, T. (2014). Antagonistic mechanisms of synbiosis between Lactobacillus plantarum CIF17AN2 and green banana starch in the proximal colon model challenged with Salmonella Typhimurium. *Anaerobe*, 28, 44–53. https://doi.org/10.1016/j.anaerobe.2014.0 5.002
- Wisnubroto, K. (2021). *Memoles Pisang Jadi Andalan Ekspor Nasional.* https://indonesia.go.id/kategori/komoditas /3194/memoles-pisang-jadi-andalanekspor-nasional

- Xu, H.-M., Huang, H.-L., Xu, J., He, J., Zhao, C., Peng, Y., Zhao, H.-L., Huang, W.-Q., Cao, C.-Y., Zhou, Y.-J., Zhou, Y.-L., & Nie, Y.-Q. (2021). Cross-Talk Between Butyric Acid and Gut Microbiota in Ulcerative Colitis Following Fecal Microbiota Transplantation. *Frontiers in Microbiology*, *12*, 658292. https://doi.org/10.3389/fmicb.2021.65829 2
- Zahid, H. F., Ranadheera, C. S., Fang, Z., & Ajlouni, S. (2021). Utilization of Mango, Apple and Banana Fruit Peels as Prebiotics and Functional Ingredients. *Agriculture*, *11*(7), 584. https://doi.org/10.3390/agriculture110705 84
- Załęski, A., Banaszkiewicz, A., & Walkowiak, J. (2013). Butyric acid in irritable bowel syndrome. *Przeglad Gastroenterologiczny*, 8(6), 350–353. https://doi.org/10.5114/pg.2013.39917
- Zarinah, Z., Anis, A. A., Napisah, H., & Shazila,
  S. (2018). Prebiotic activity score of breadfruit resistant starch (Artocarpus altilis), Breadfruit flour, and inulin during in-vitro fermentation by pure cultures (Lactobacillus plantarum, and Bifidobacterium bifidum). *Journal of Agrobiotechnology*, 9(1S), 122–131.
- Zhao, X., Zhong, X., Liu, X., Wang, X., & Gao, X. (2021). Therapeutic and Improving Function of Lactobacilli in the Prevention and Treatment of Cardiovascular-Related Diseases: A Novel Perspective From Gut Microbiota . In *Frontiers in Nutrition* (Vol. 8). https://www.frontiersin.org/articles/10.33

https://www.frontiersin.org/articles/10.33 89/fnut.2021.693412